



Size distribution of *Haliotis asinina* and abundance of periphyton in tapulaga waters

Ermayanti Ishak^{1,*}, Muhammad Taswin Munier¹, Muhammad Trial²

¹ Aquatic Resources Management, Faculty of Fisheries and Marine Sciences, Halu Oleo University, Kendari, Indonesia.

² Marine Science, Faculty of Fisheries and Marine Sciences, Halu Oleo University, Kendari, Indonesia.

ARTICLE INFO

ABSTRACT

Keywords:

H. asinina
Macroalgae
Periphyton
Tapulaga

The Tapulaga waters, located in Soropia, Southeast Sulawesi, are among the coastal waters with significant potential for the seven-eyed abalone (*Haliotis asinina*). This species is a herbivorous gastropod that primarily consumes natural food sources such as periphyton and macroalgae. This study aims to determine the size distribution of *H. asinina* and analyze the presence of periphyton on seagrass leaves and macroalgal thalli. Sampling was conducted over six months in the Tapulaga waters of Soropia, Konawe Regency, Southeast Sulawesi, using a simple random sampling technique. The collected data were analyzed using descriptive statistical methods. The results indicate that the highest percentage of size distribution was observed in the range of 37.5–44.5 mm, comprising 34% of the total population. The population was categorized into two size groups: juveniles (≤ 49 mm) and adults (≥ 50 mm). Juveniles accounted for 75% of the total *H. asinina* population, while adults comprised 25%. A total of 11 periphyton species from three taxonomic classes Bacillariophyceae, Cyanophyceae, and Trebouxiophyceae—were identified on the seagrass species *Enhalus acoroides* and the macroalgae *Gracilaria salicornia*, *Padina* sp., *Halimeda* sp., *Galaxaura* sp., and *Gelidium* sp. The identified periphyton species included *Aulacoseira* sp., *Nitzschia* sp., *Synedra* sp., *Diatoma* sp., *Fragilaria* sp., *Navicula* sp., *Pinnularia* sp., *Melosira* sp., *Lyngbya* sp., *Planctonema* sp., and *Cocconeis* sp. Six of these periphyton species were present on both seagrass and macroalgal thalli. *Synedra* sp. was the most dominant species, exhibiting the highest abundance at 88,876 individuals/cm². The presence of periphyton as a natural food source attached to seagrass leaves and macroalgal thalli plays a crucial role in supporting the juvenile *H. asinina* population.

DOI: 10.13170/depik.14.1.42737

Introduction

Haliotis asinina, commonly known as the "mata tujuh" (seven-eyed) abalone (Dharma, 2005; Setyono, 2004; Ishak *et al.*, 2020), is a gastropod species of high economic value, particularly in the fishing and aquaculture industries (Setyono, 2009a). This species is widely distributed across the eastern waters of Indonesia (Gallardo & Salayo, 2003; Setyono, 2009b), including the Tapulaga waters in Soropia, Southeast Sulawesi (Ishak *et al.*, 2020).

The waters of Tapulaga Village serve as one of the habitats for *H. asinina*, alongside other locations in the Soropia District, Konawe Regency. This area is characterized by a seagrass ecosystem, which is exposed during low tide and submerged during high tide, making it part of the intertidal zone (Yulianda *et al.*, 2013). Various species of macroalgae also inhabit

this environment (Handayani, 2021), creating an ideal habitat for *H. asinina*.

Understanding the size distribution of populations is crucial for ecological studies and conservation efforts, as it provides insights into demographic structure, population dynamics, and overall ecosystem health. Population size distribution is influenced by multiple environmental factors, including food availability, predation, and habitat conditions (Ardiyansyah, 2018). Periphyton, or benthic diatoms, serve as a primary natural food source for abalones and play a significant role in their ecology (Nasrullah *et al.*, 2016). Periphyton consists of a community of organisms that attach to various submerged surfaces in aquatic environments, such as rocks, bamboo substrates (Sibarani *et al.*, 2020), cement plates, nets, plastics (Nasrullah *et al.*, 2016; Zain *et al.*, 2023), and seagrass leaves (Pabesak, 2004).

* Corresponding author.

Email address: ermayanti.ishak@uho.ac.id

The abundance and composition of periphyton directly influence the survival rates of *H. asinina* larvae (Jarayabrand & Paphavasit, 1996; Xing *et al.*, 2008; Nasrullah *et al.*, 2016).

This study focuses on analyzing the size distribution of *H. asinina* and the abundance of periphyton to better understand the relationship between population structure and habitat conditions. Examining size distribution provides valuable insights into individual size variation, growth patterns, and overall population dynamics, which are essential for assessing reproductive rates, maturity levels, and ecological pressures. Additionally, investigating periphyton abundance helps assess aquatic ecosystem health, as periphyton is sensitive to environmental changes and can serve as a biological indicator of water quality, nutrient levels, pollution, and marine productivity.

Understanding the relationship between *H. asinina* size distribution and periphyton abundance is expected to provide valuable information for the effective management and conservation of this species. Furthermore, the findings of this study may contribute to the development of sustainable abalone aquaculture and enhance conservation efforts for the aquatic ecosystems that support *H. asinina*.

Materials and Methods

Study Location and Duration

Sampling was conducted monthly over a six-month period in the seagrass ecosystem of the Tapulaga waters, Soropia District, Southeast Sulawesi (Figure 1)

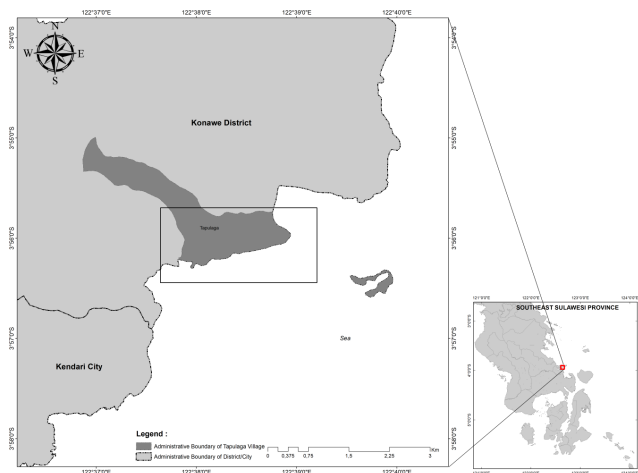


Figure 1. Map of the Research Site

Data Analysis

Research Tools and Materials

The equipment used in this study included digital calipers, rulers, microscopes, sample bottles, ethanol,

brushes, trays, and periphyton identification guides (Prescott, 1970; Clark, 1981).

Data Collection Procedure

A simple random sampling technique was employed, and data were analyzed using descriptive statistics. The shell length of *H. asinina* was measured using a digital caliper with an accuracy of 0.1 mm. Periphyton samples were collected from the leaves of *Enhalus acoroides* seagrass and the thalli of various macroalgae species present at the sampling site. The leaves and thalli were standardized to the same length using a ruler and cut to approximately 10 cm² before being placed into sample bottles containing 4% formalin.

Periphyton (microalgae) observations were conducted using a light microscope at 40× magnification with a Sedgewick-Rafter Counting Cell (SRC) slide. Before observation, the periphyton was detached from the leaves and thalli by scraping them with a brush. The collected periphyton was then transferred into sample bottles containing 50 mL of water and preserved in 4% formalin. Periphyton identification was based on morphological characteristics following standard references (Yamaji, 1979; Biggs and Kilroy, 2000; Andrzej *et al.*, 2000).

Data analysis

The shell length data were analyzed using Sturges' formula (1926) as cited in Lempoy (2020) to determine the frequency distribution of the size classes.

$$k = 1 + 3,3 \log n$$

Where: k = number of classes, n = number of data points. The class interval was calculated using the following formula:

$$C = \frac{X_n - X_1}{k}$$

Where: C = class interval, X_n = highest data value; X₁ = lowest data value, k = number of classes.

Periphyton abundance was calculated using the formula by Ameilda *et al.* (2016):

$$N = \frac{n \times At \times Vt}{AC \times Vs \times As}$$

Where: N = periphyton abundance (ind/cm²), n = number of observed periphyton individuals, At = cover glass area (22 x 22 mm²), Vt = total sample volume in the sample bottle (25 ml), Ac = microscope field area (1.036 mm²); Vs = volume of one drop of sample under the cover glass (0.5 ml); As = scraped substrate surface area (5 x 5 cm²).

Results

Size Distribution of *H. asinina*

Based on shell length measurements of *H. asinina*, the highest frequency (34%) was observed in the size class of 37.5–44.5 mm, which falls within the juvenile category (Figure 2).

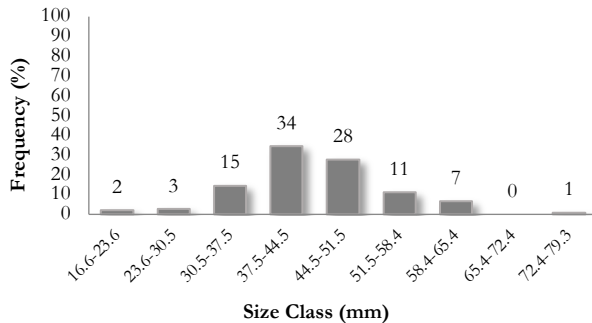


Figure 2. Distribution of frequency of abalón shell's size in several size classes

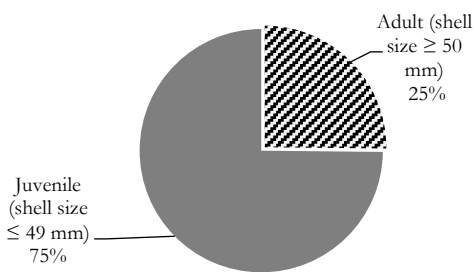


Figure 3. Percentage of Size Groups of Abalón *H. asinina* found in Tapulaga Waters

Analysis of shell length data from a total of 151 *H. asinina* individuals collected from the Tapulaga waters revealed that 25% of the population were adults (≥ 50 mm in shell length), while 75% were juveniles (≤ 49 mm in shell length). This indicates that the majority of the population consists of juveniles.

Periphyton Abundance

The types of periphyton and macroalgae identified in the Tapulaga waters are presented in Figure 3.

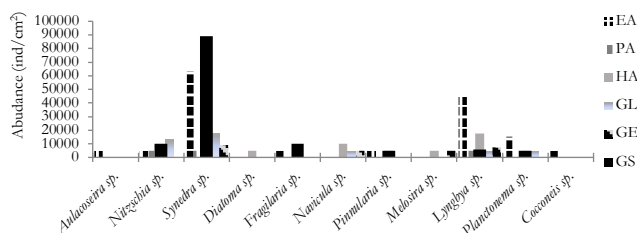


Figure 4. The types of periphyton and macroalgae identified in the Tapulaga waters

As shown in Figure 4, a total of 11 periphyton species were recorded in the Tapulaga waters, including *Aulacoseira* sp., *Nitzschia* sp., *Synedra* sp., *Diatoma* sp., *Fragilaria* sp., *Navicula* sp., *Pinnularia* sp., *Melosira* sp., *Lyngbya* sp., *Planctonema* sp., and *Cocconeis* sp. These species were identified on the leaves of the seagrass *Enhalus acoroides* and the thalli of several macroalgae species, including *Gracilaria salicornia*, *Padina* sp., *Halimeda* sp., *Galaxaura* sp., and *Gelidium* sp.

The same periphyton species were found on both seagrass leaves and macroalgal thalli. The abundance of periphyton on *E. acoroides* leaves and macroalgal thalli in the Tapulaga waters ranged from 5,000 to 88,876 individuals/cm². Among the recorded species, *Synedra* sp., from the class Bacillariophyceae, was the most dominant and exhibited the highest abundance across all observations on seagrass leaves and macroalgal thalli.

Discussion

The analysis indicates that the Tapulaga waters exhibit a high frequency of *Haliotis asinina* within the shell length size class of 37.5–44.5 mm, representing 34% of the population. Among the 151 *H. asinina* individuals sampled, 75% had shell sizes ≤ 49 mm, classifying them as juveniles, while 25% had shell sizes ≥ 50 mm, categorizing them as adults. The juvenile category is further subdivided into early juveniles and juveniles, whereas the adult category comprises subadults and adults. Early juveniles typically have a shell length between 5–10 mm (Franco *et al.*, 2022), while juveniles range from 24.3–25.3 mm (Jwa and Hong, 2023) or approximately 20.2 mm (Nivelais, 2023). Small abalone generally measure around 15.8 ± 1.6 mm (Rizzo *et al.*, 2024). Juvenile abalone exhibit shell lengths between 21 and 38 mm (Bullon *et al.*, 2023). Abalone seeds range from 2.0–3.5 cm (WWF Indonesia and Rusdi, 2015). Small-sized abalone are classified as those measuring less than 40 mm (Ishak *et al.*, 2019). Ripe females typically have shell lengths between 58.1–69 mm (Capinpin *et al.*, 1998).

Adult abalone generally measure over 50 mm (Ishak *et al.*, 2020; WWF Indonesia and Rusdi, 2015; Xiaobing *et al.*, 2010). The minimum harvest size for green abalone (*Haliotis fulgens*) is 150 mm (Bauer *et al.*, 2024). The onset of gonadal maturity in *H. asinina* occurs at a shell length of 64 mm (Hadijah, 2017).

The high percentage of juveniles observed in this study (Figure 1) is closely related to the availability of natural food sources. Juvenile abalone primarily consume periphyton or benthic diatoms from the microalgae group, which are found attached to

seagrass leaves and macroalgal thalli. Observations identified 11 types of periphyton on seagrass leaves and macroalgal thalli, including *Aulacoseira* sp., *Nitzschia* sp., *Synedra* sp., *Diatoma* sp., *Fragilaria* sp., *Navicula* sp., *Pinnularia* sp., *Melosira* sp., *Lynghya* sp., *Planctonema* sp., and *Cocconeis* sp. Six periphyton species were found on both seagrass and macroalgae: *Nitzschia* sp., *Synedra* sp., *Fragilaria* sp., *Pinnularia* sp., *Lynghya* sp., and *Planctonema* sp. Among these, *Nitzschia* sp. serves as a suitable natural food source for early juvenile *H. asinina*. Other important periphyton species for juvenile *H. asinina* include *Cocconeis* sp., *Nitzschia* sp., *Amphora* sp., and *Navicula ramosissima* (Villa-Franco *et al.*, 2022).

The periphyton species preferred by juvenile abalone include *Nitzschia* sp., *Navicula* sp., and *Cocconeis* sp. (Imai, 1982). Additionally, *Navicula mollis*, *Stauroneis* sp., *N. ramosissima*, *Pleurosigma* sp., and *Cocconeis* sp. are benthic diatoms that stimulate metamorphosis and growth, with cell lengths ranging from $13.64 \pm 0.29 \mu\text{m}$ to $26.54 \pm 0.45 \mu\text{m}$ (Capinpin, 2015). *Cocconeis* sp., *Nitzschia* sp., *Amphora* sp., and *Navicula ramosissima* are considered suitable live feeds for early juvenile *H. asinina* (Villa-Franco *et al.*, 2022). The most essential natural food sources during the early juvenile stage are *Nitzschia* sp. and *Navicula* sp., as they provide key nutrients for early abalone growth (Setyono, 2009a). Abalone larvae primarily consume benthic diatoms, while adults feed on seaweed or macroalgae (Nurfajrie *et al.*, 2014).

The abundance of *Synedra* sp. ranged from 5,000 to 88,876 individuals/cm². The high abundance of *Synedra* sp. observed on all surfaces of seagrass leaves and macroalgal thalli suggests potential environmental pollution; however, further studies are required to determine the pollution category. The highest periphyton abundance of *Synedra* sp. was recorded on *Enhalus acoroides* leaves (63,333 individuals/cm²) and on the macroalgae *Gracilaria salicornia* thalli (88,876 individuals/cm²). *Synedra* sp., a periphyton species from the class Bacillariophyceae, exhibited the highest abundance among all identified periphyton species. This species is known for its ability to survive in unfavorable environmental conditions (Conradie, 2008) and can thrive in low-nitrogen and phosphate conditions (Venter, 2003). It is also recognized as a bioindicator for water quality assessment and is often associated with pollution in aquatic ecosystems (Rangpan, 2008; Isti'annah *et al.*, 2015).

In the Tapulaga waters, adult *Halotis asinina* were present, although in smaller numbers (25%). Adult *H. asinina* exhibit greater selectivity in their diet compared to juveniles, primarily consuming

macroalgae (Permana *et al.*, 2017). Both adults and larvae have been reported to favor a combination of benthic algae (Hadijah *et al.*, 2017), and even juvenile *H. squamata* consume macroalgae (Damayanti *et al.*, 2018). Adults show a preference for brown macroalgae over green algae (Imai, 1982). Other studies indicate that red algae, particularly *Gracilaria* sp., promote growth and gonad maturation in *H. asinina* (Singhagraiwan and Doi, 1993; Capinpin and Corre, 1996; Priyambodo *et al.*, 2005; Setyono, 2006; Hadijah *et al.*, 2014). This type of algae has been shown to support optimal growth in abalone (Susanto *et al.*, 2008; Rusdi *et al.*, 2010) and is commonly used as the primary feed in floating cage aquaculture (WWF Indonesia and Rusdi, 2015). Other macroalgae species preferred by abalone include *Gracilaria verrucosa*, *Ulva* sp., and *Kappaphycus* sp. (Setyono, 2009a).

The low percentage of adult *H. asinina* in Tapulaga may be influenced by the types of macroalgae available, which may be less suitable for their dietary preferences. The macroalgae species found in Tapulaga include *Gracilaria salicornia*, *Padina minor*, *Halimeda* sp., *Galaxaura* sp., and *Gelidium* sp. Among these, *Gracilaria* sp. is particularly favored by abalone due to its relatively soft thallus texture (Chen, 1984; Rusdi *et al.*, 2010), smaller branches (Hadijah *et al.*, 2017), and slightly thinner cell walls with pseudophylls (Erniati *et al.*, 2022). Additionally, *Gracilaria* sp. serves as a natural food source that enhances both growth and survival in abalone (Reyes and Fermin, 2003; Naidoo *et al.*, 2006; Qi *et al.*, 2010). This macroalgae species also contains the probiotic Alg 3.1, which enhances nutrient availability in the digestive system of *H. asinina* (Faturrahman, 2013). In contrast, *G. salicornia* is characterized by a large and dense thallus (Tuiyo, 2014), while *Ulva* sp. has a soft texture that is easily digestible by juvenile abalone (Chen, 1984). *Halimeda* sp., in contrast, is green and has a stiff thallus (Erniati *et al.*, 2022).

Several factors influence the selection of macroalgae as a food source for abalone, including algal morphology, texture, hardness, metabolite composition, and nutritional content (Shepherd and Steinberg, 1992; Masita *et al.*, 2016; Akbar and Hasan, 2024). Abalone consumes algae in quantities ranging from 10% to 30% of their body weight daily (Chen, 1989). As herbivorous marine gastropods, abalone feed on both microalgae and macroalgae, including red, green, and brown algae (Faturrahman *et al.*, 2015). In their natural habitat, most adult abalone are sedentary, capturing floating macroalgae from the water column (Allen *et al.*, 2006). They also consume benthic microalgae when macroalgae resources are

scarce, such as in tropical coral reef environments (Sawatpeera *et al.*, 1998). In tropical regions, abalone consumes various macroalgae species, including *Gracilaria* sp., *Laurencia obtusa*, *Ulva* sp., *Hypnea asper*, and *Kappaphycus alvarezii* (Setyono, 2009a).

The texture of macroalgal thalli plays a crucial role in abalone food selection, along with its chemical composition and nutritional value. *Gracilaria verrucosa*, for example, is rich in protein and fiber and contains essential amino acids such as L-threonine, L-glycine, L-leucine, and L-alanine (Ma'ruf *et al.*, 2013).

Conclusion

The size distribution of *Haliotis asinina* in Tapulaga indicates that juveniles constitute the majority of the population. This high abundance of juvenile abalone is likely supported by the availability of natural food sources, particularly microalgae, including periphyton and benthic diatoms, which thrive on seagrass leaves and macroalgae thalli. Among the periphyton species identified, *Synedra* sp. was the most dominant. The high prevalence of this species may indicate potential environmental pollution; however, further research is required to assess the extent and nature of the pollution.

References

- Akbar, S. A., M. Hasan. 2024. Evaluation of bioactive composition and phytochemical profile of macroalgae *Gracilaria edulis* and *Acanthophora spicifera* from the Banda Aceh Coast, Indonesia. *Science & Technology Asia*, 29(1): 194–207.
- Ameilda, C. H., I. Dewiyanti, C. Octavina. 2016. Community structure of periphyton on the macroalga *Ulva lactuca* in coastal waters of Ulee Lheue Beach, Banda Aceh. *Journal of Student Scientific and Maritime Unsyiah*, 1(3): 337–347.
- Allen, V. J., I. D. Marsden, N. I. C. Ragg, S. Gieseg. 2006. The effects of tactile stimulants on feeding, growth, behaviour, and meat quality of cultured blackfoot abalone, *Haliotis iris*. *Aquaculture*, 257(1–4): 294–308.
- Andrzej, W., H. Lange-Bertalot, D. Metzeltin. 2000. Diatom flora of marine coasts I. *Iconographia Diatomologica*, 7: 1–925.
- Ardiyansyah, F. 2018. Distribution patterns and composition of gastropods at the Kukur TN Alas Purwo resort. *Journal of Biology and Biology Learning*, 3(2): 139–151.
- Bauer, J., J. Segovia-Rendon, J. Lorda, A. Abadia-Cardoso, L. Malpica-Cruz, P. Alvarado-Graef, R. Searcy-Bernall, L. Vazquez-Vera, R. Beas-Luna. 2024. Short-term effects of community-based marine reserves on green abalone, as revealed by population studies. *Scientific Reports*, 14: 955.
- Biggs, B. J. F., C. Kilroy. 2000. *Stream Periphyton Monitoring Manual*. NIWA, New Zealand.
- Bullon, N., A. Setfoddin, S. M. Dezfooli, T. Young, A. C. Alfaro. 2023. Nutritional and metabolomics changes of juvenile farmed abalone (*Haliotis iris*) in New Zealand. *Aquaculture Research*, 1–17.
- Capinpin, E. C., K. Corre. 1996. Growth rate of the Philippine abalone, *Haliotis asinina* fed an artificial diet and macroalgae. *Aquaculture*, 144: 81–89.
- Capinpin, E. C., V. C. Encena, N. C. Bayona. 1998. Studies on reproductive biology of the Donkey's ear abalone, *Haliotis asinina* Linne. *Aquaculture*, 166: 141–150.
- Capinpin, E. C. 2015. Settlement of the tropical abalone *Haliotis asinina* on different diatoms. *International Journal of Fauna and Biological Studies*, 2(1): 30–34.
- Chen, H. C. 1989. Farming the small abalone, *Haliotis diversicolor supertexta* in Taiwan. In: Hahn, K. O. (Ed.), *Handbook of Culture of Abalone and Other Marine Gastropods*. CRC Press, Boca Raton, FL. pp. 265–283.
- Chen, H. C. 1984. Recent innovation in cultivation of edible molluscs in Taiwan, with special reference to the small abalone *Haliotis diversicolor* and the hard clam *Meretrix lusoria*. *Aquaculture*, 39: 11–29.
- Conradie, K. R., S. Du Plessis, A. Venter. 2008. School of environmental sciences and development: botany. *South African Journal of Botany*, 74: 101–110.
- Damayanti, D., D. S. Yusup, I. Rusdi. 2018. The effect of feeding some macroalgae (*Ulva* sp., *Gracilaria* sp., *Halymenia* sp.) on the growth of abalone *Haliotis squamata*. *Journal of Metamorphosis*, 5(2): 189–197.
- Dharma, B. 2005. *Recent & fossil Indonesian shells*. ConchBooks, Jakarta. 424 pp.
- Erniati, Erlangga, Y. Andika. 2022. *Seaweed (Aceh aquatic)*. KBM Indonesia, Jawa Timur, Indonesia. 73 pp.
- Faturrahman. 2013. Improved of *Gracilaria* sp. digestibility on abalone (*Haliotis asinina*) by giving agar-degrading probiotic. *UNRAM Research Journal*, 17(2): 136–141.
- Faturrahman, A. Meryandini, M. Z. Junior, I. Rusmana. 2015. The role of agarolytic bacteria in enhancing physiological function for digestive system of abalone (*Haliotis asinina*). *Journal of Applied Environmental Biology Science*, 5(5): 495–620.
- Gallardo, W. G., N. D. Salayo. 2003. Abalone culture: a new business opportunity. *SEAFDEC Asian Aquaculture*, 25(3): 25–28.
- Hadijah, M. Rahmiyanti, S. Mulyani. 2014. Effect of *Gracilaria* sp. feed dose on the growth of *Haliotis squamata* abalone seeds. *Proceedings of the First National Symposium on Maritime Affairs and Fisheries 2014*. Faculty of Marine and Fishery Sciences, Hasanuddin University, Makassar. ISBN 976-602-71759-0-7.
- Hadijah, A. Tuwo, M. Litaay, E. Indrawati. 2017. The reproductive aspect of tropical abalone (*Haliotis asinina* L.) in the waters of Tanakeke Islands at South Sulawesi. *Aquatic Science and Technology*, 1(2): 30–43.
- Hadijah. 2017. *Get to know tropical abalone: biology and ecology*. Sah Media, Makassar, Indonesia. 150 pp.
- Handayani, T. 2021. Diversity of macroalgae in the waters of Kendari Bay and its surroundings, Southeast Sulawesi. *Oceanology and Limnology in Indonesia*, 6(1): 55–69.
- Imai, T. 1982. *Aquaculture in shallow seas: progress in shallow sea culture*. VI. *Artificial Culture of Shellfish (Artificial Seeding of Abalone)*. A.A. Balkema, Rotterdam. pp. 374–606. ISBN-13: 9789061910220.
- Ishak, E., I. Setyobudiandi, F. Yulianda, M. Boer. 2019. Sex ratio and minimum size at sexual maturity of *Haliotis asinina* in the seagrass vegetation of Tapulaga, Southeast Sulawesi, Indonesia. *AAAL Bioflux*, 12(5): 1635–1642.
- Ishak, E., I. Setyobudiandi, F. Yulianda, M. Boer, Bahtiar. 2020. Effects of habitat type diversity on population structure and morphometrics of the abalone *Haliotis asinina* Linnaeus 1758. *Journal of Tropical Biology*, 20(1): 29–39.
- Isti'anah, D., M. F. Huda, A. N. Laily. 2015. *Synedra* sp. as microalgae found in the Besuki Porong River, Sidoarjo, East Java. *Bioeducation*, 8(1): 57–59.
- Jarayabhand, P., N. Paphavasit. 1996. A review of the culture of tropical abalone with special reference to Thailand. *Aquaculture*, 140: 159–168.
- Jwa, M. S., C. Y. Hong. 2023. Abiotic stress tolerance of juvenile small abalone *Haliotis diversicolor aquatilis* (Obunjagi) to gamma irradiation. *Journal of Coastal Research*, 39(1): 55–62.
- Lempoy, R., A. B. Rondonuwu, N. E. Bataragoa. 2020. Size and length, weight of fish and condition factors of the Banggao dragonfish *Pterapogon kauderni* Koumans, 1933 in the Lebeh Strait, North Sulawesi. *Platax Scientific Journal*, 8(1): 30–36.
- Ma'ruf, W. F., R. Ibrahim, E. N. Dewi, E. Susanto, U. Amalia. 2013. Profile of seaweed *Caulerpa racemosa* and *Gracilaria verrucosa* as edible food. *Fisheries Science Journal*, 9(1): 68–74.
- Masita, I. J. Effendy, A. B. Patadjai. 2016. Feed consumption and gonad maturation of abalone (*Haliotis asinina*) cultured at integrated

- multi-trophic aquaculture (IMTA) system using different natural feed. *Media Akuatika*, 1(1): 55–61.
- Naidoo, K., K. G. Maneveldt, J. Ruck, J. J. Bolton. 2006. A comparison of various seaweed-based diets and formulated feed on growth rate of abalone in a land-based aquaculture system. *Journal of Applied Phycology*, 18: 437–443.
- Nahrullah, M. Idris, I. J. Effendy. 2016. The utilization of different substrates for the production of early juvenile (50 days old) abalone (*Haliotis asinina*). *Media Akuatika*, 1(3): 182–189.
- Nivelais, L., A. Levallois, O. Basuyaux, K. Costil, J. M. Lebel, S. Larsonneur, G. Gulchard, A. Serpentine, C. Caplat. 2023. Effect on growth of juvenile *Haliotis tuberculata* under chronic exposition to metals released from the dissolution of an aluminium-based galvanic anode. *Archives of Environmental Contamination and Toxicology*, 84(1): 1–13.
- Nurfajrie, Suminto, S. Rejeki. 2014. Utilization of various types of microalgae for the growth of abalone (*Haliotis squamata*) in grow-out cultivation. *Journal of Aquaculture Management and Technology*, 3(4): 142–150.
- Pabesak, E. 2004. Species composition and density of benthic diatoms in Purirano coastal waters. [Thesis]. Faculty of Fisheries and Marine Sciences, Halu Oleo University, Kendari. 50 p.
- Permana, G. N., F. H. Khotimah, B. Susanto, I. Rusdi, Haryanti. 2017. Performance of growth and reproduction of abalone *Haliotis squamata* Reeve (1846) third generation. *Journal of Aquaculture Research*, 12(3): 197–202.
- Priyambodo, B., Y. Sofyan, Jaya Suastika, I. B. M. 2005. Production of abalone oyster seeds (*Haliotis asinina*) at the Loka of Lombok Marine Aquaculture. Proceedings of the Annual National Seminar on Fisheries and Marine Research Results, UGM, Yogyakarta: 5.
- Qi, Z., H. Liu, B. Li, Y. Mao, Z. Jiang, J. Zhang, J. Fang. 2010. Suitability of two seaweeds, *Gracilaria lemaneiformis* and *Sargassum pallidum*, as feed for the abalone *Haliotis discus hannai* Ino. *Aquaculture*, 300(1–4): 189–193.
- Rangan, V. 2008. Effects of water quality on periphyton in the Pattani River, Yala Municipality, Thailand. [Thesis] Universiti Sains Malaysia, Malaysia.
- Reyes, O. S., A. C. Fermin. 2003. Terrestrial leaf meals or freshwater aquatic fern as potential feed ingredients for farmed abalone, *Haliotis asinina* (Linnaeus 1758). *Aquaculture Research*, 34(8): 593–599.
- Rizzo, N. R., S. B. Beckert, S. E. Boles, J. A. Gross. 2024. Temperature-induced variations in dulse (*Devaleraea mollis*) nutrition provide indirect benefits on juvenile red abalone (*Haliotis rufescens*) growth. *Frontiers in Marine Science*, 11: 1336793.
- Rusdi, I., R. Rahmawati, B. Susanto, I. N. Adiasmara. 2010. Maturation of gonads of *Haliotis squamata* abalone broodstock through feed management. *Journal of Aquaculture Research*, 5(3): 383–391.
- Sarbini, R., Y. Nugraha, H. Kuslani. 2015. Sampling technique and observation of periphyton abundance in seagrass ecosystems, Karimun Jawa Islands, Central Java. *Resources and Capture Research Engineering Bulletin*, 13(2): 91–96.
- Sawatpeera, S., E. S. Upatham, M. Kruatrachue, V. Ingsrisawang, T. Singhagraiwan, Y. P. Chitramvong, K. Parkpoomkamol. 1998. Determination of gut contents of Thai abalone *Haliotis asinina* Linnaeus. *Journal of Shellfish Research*, 17(3): 765–769.
- Setyono, D. E. D. 2004. Abalone (*Haliotis asinina* L.): Prospective species for aquaculture in Indonesia. *Oseana*, 29(2): 25–30.
- Setyono, D. E. D. 2006. Induction spawning for the tropical abalone (*Haliotis asinina*) in the laboratory. *Indonesian Aquaculture Journal*, 1(1): 17–27.
- Setyono, D. E. D. 2009a. Abalone: Biology and reproduction. UPT Loka Marine Bio Industry Development, Indonesian Institute of Sciences (LIPI) Press, Mataram. 92 p.
- Setyono, D. E. D. 2009b. Abalone: Seeding technology. Association of Indonesian Oceanology Scholars (ISOI), Jakarta Utara. 14 p.
- Shepherd, S. P., P. D. Steinberg. 1992. Food preference of three Australian abalone. In: *Abalone of the world: Biology, fisheries and culture*. Proceedings of the 1st International Symposium on Abalone Fishing.
- Sibarani, L. B. G., T. Dahril, A. H. Simarmata. 2020. Types and abundance of periphyton with bamboo substrates in the puddles of Batu Bersurat Village, Kampar Regency, Riau. *Journal of Aquatic Resources and Environment*, 1(1): 81–92.
- Singhagraiwan, T., M. Doi. 1993. Seed production and culture of a tropical abalone, *Haliotis asinina* Linne. Research Project of Fishery Resource Development in the Kingdom of Thailand, Bangkok, Thailand.
- Tuiyo, R. 2014. Identification of red algae (*Gracilaria* sp.) in Gorontalo Province. *Scientific Journal*, 7(4): 379–383.
- Venter, A. A., A. J. H. Jordaan, J. Pieterse. 2003. *Oscillatoria simplicissima*: A taxonomical study. School of Environmental Sciences and Development: Botany. *Journal Water SA*, 29(1).
- Villa-Franco, A. U., M. R. D. Pena, M. F. J. Nievales. 2022. Grazing periodicity, grazing rate, feeding preference, and gut examination of early juveniles of abalone *Haliotis asinina* fed five benthic diatom species. *Aquaculture International*, 30: 2343–2364.
- WWF Indonesia, I. Rusdi. 2015. Abalone cultivation (*Haliotis* sp.) floating karamba system. 1st Edition.
- Xing, R. I., C. Wang, X. B. Chang, Y. Chang. 2008. Settlement, growth and survival of abalone, *Haliotis discus hannai*, in response to eight monospecific benthic diatoms. *Journal of Applied Phycology*, 20: 47–53.
- Yamaji, I. 1979. Illustration of marine plankton of Japan. Vaikiska Publication, Japan. 593 p.
- Yulianda, F., M. S. Yusuf, W. Prayogo. 2013. Zonation and density of intertidal communities at coastal area of Batu Hijau, Sumbawa. *Journal of Tropical Marine Science and Technology*, 5(2): 409–416.
- Zain, Y. G., M. Junaidi, L. F. Mulyani. 2023. The effect of different substrates on the growth and survival of abalone (*Haliotis squamata*). *Laot Journal of Marine Science*, 5(2): 204–218.

How to cite this paper:

Ishak, E., M. T. Munier, M. Trial. 2025. Size distribution of *Haliotis asinina* and abundance of periphyton in Tapulaga waters. *Depik: Jurnal Ilmu-Ilmu Perairan, Pesisir dan Perikanan*, 14(1): 12–17.